

NUCLEOTIDE SEQUENCES CODING FOR THE sigC GENE

[0001] BACKGROUND OF THE INVENTION

The subject of the present invention are nucleotide sequences of coryneform bacteria coding for the sigC gene and a process for the enzymatic production of amino acids using bacteria in which the sigC gene is enhanced. All references cited herein are expressly incorporated by reference. Incorporation by reference is also designated by the term "I.B.R." following any citation.

[0002] L-amino acids are used in human medicine and in the pharmaceutical industry, in the foodstuffs industry and, most especially, in animal nutrition.

[0003] It is known that amino acids can be produced by fermentation of strains of coryneform bacteria, in particular *Corynebacterium glutamicum*. On account of the great importance of amino acids efforts are constantly being made to improve the production processes. Process improvements may involve fermentation technology measures such as for example stirring and provision of oxygen, or the composition of the nutrient media, such as for example the sugar concentration during the fermentation, or the working-up to the product form by for example ion exchange chromatography or the intrinsic performance properties of the microorganism itself.

[0004] In order to improve the performance properties of these microorganisms methods involving mutagenesis, selection and mutant selection are employed. In this way strains are obtained that are resistant to antimetabolites or are auxotrophic for regulatory important metabolites, and that produce amino acids.

[0005] For some years methods of recombinant DNA technology have also been used to improve L-amino acid-producing strains of *corynebacterium*, by amplifying individual amino

acid biosynthesis genes and investigating the effect on amino acid production.

[0006] The invention provides new techniques for the improved enzymatic production of amino acids.

[0007] BRIEF SUMMARY OF THE INVENTION

When L-amino acids or amino acids are mentioned hereinafter, it is understood that this refers to one or more amino acids including their salts, selected from the group L-asparagine, L-threonine, L-serine, L-glutamate, L-glycine, L-alanine, L-cysteine, L-valine, L-methionine, L-isoleucine, L-leucine, L-tyrosine, L-phenylalanine, L-histidine, L-lysine, L-tryptophan and L-arginine. Lysine is particularly preferred.

[0008] The present invention provides an isolated polynucleotide from coryneform bacteria containing a polynucleotide sequence coding for the sigC gene, selected from the group

- a) polynucleotide that is at least 70% identical to a polynucleotide coding for a polypeptide that contains the amino acid sequence of SEQ ID No. 2,
- b) polynucleotide coding for a polypeptide that contains an amino acid sequence that is at least 70% identical to the amino acid sequence of SEQ ID No. 2,
- c) polynucleotide that is complementary to the polynucleotides of a) or b), and
- d) polynucleotide containing at least 15 successive nucleotides of the polynucleotide sequence of a), b) or c),

the polypeptide preferably having the activity of the sigma factor C.

[0009] The present invention also provides the aforementioned polynucleotide, which is preferably a replicable DNA containing:

- (i) the nucleotide sequence shown in SEQ ID No. 1, or
- (ii) at least one sequence that corresponds to the sequence (i) within the region of degeneracy of the genetic code, or
- (iii) at least one sequence that hybridizes with the sequence that is complementary to the sequence (i) or (ii), and optionally
- (iv) functionally neutral sense mutations in (i).

[0010] The invention furthermore provides

a replicable polynucleotide, in particular DNA, containing the nucleotide sequence as shown in SEQ ID No. 1;

a polynucleotide coding for a polypeptide that contains the amino acid sequence as shown in SEQ ID No. 2;

a vector containing the polynucleotide according to the invention, in particular a shuttle vector or plasmid vector, and

coryneform bacteria that contain the vector or in which the sigC gene is enhanced.

[0011] The present invention moreover provides polynucleotides that consist substantially of a polynucleotide sequence that can be obtained by screening by means of hybridization of a corresponding gene library of a coryneform bacterium that contains the complete gene or parts thereof, with a probe that contains the sequence of the polynucleotide of the invention according to SEQ ID No. 1 or a fragment thereof, and isolation of the aforementioned polynucleotide sequence.

[0012] BRIEF DESCRIPTION OF THE FIGURES

Fig. 1: Map of the plasmid pEC-XK99E

Fig. 2: Map of the plasmid pEC-XK99EsigCb2ex

The abbreviations and acronyms used have the following meanings:

Kan:	Kanamycin resistance gene aph(3')-IIa from Escherichia coli
HindIII	Cleavage site of the restriction enzyme HindIII
XbaI	Cleavage site of the restriction enzyme XbaI
KpnI	Cleavage site of the restriction enzyme KpnI
Ptrc	trc promoter
T1	Termination region T1
T2	Termination region T2
Per	Replication effector per
Rep	Replication region rep of the plasmid pGA1
LacIq	lacIq repressor of the lac operon of Escherichia coli
SigC	Cloned sigC gene

[0013] DETAILED DESCRIPTION OF THE INVENTION

Polynucleotides that contain the sequences according to the invention are suitable as hybridization probes for RNA, cDNA and DNA in order to isolate nucleic acids or polynucleotides or genes in their full length that code for the sigma factor C, or to isolate such nucleic acids or

polynucleotides or genes that have a high sequence similarity to that of the sigC genes. They are also suitable for incorporation in so-called "arrays", "micro arrays" or "DNA chips" in order to detect and determine the corresponding polynucleotides.

[0014] Polynucleotides that contain the sequences according to the invention are furthermore suitable as primers with the aid of which, and by employing the polymerase chain reaction (PCR), DNA of genes can be produced that code for the sigma factor C.

[0015] Such oligonucleotides serving as probes or primers contain at least 25, 26, 27, 28, 29 or 30, preferably at least 20, 21, 22, 23 or 24, and most particularly preferably at least 15, 16, 17, 18 or 19 successive nucleotides. Also suitable are oligonucleotides with a length of at least 31, 32, 33, 34, 35, 36, 37, 38, 39 or 40, or at least 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 nucleotides. Also suitable if necessary are oligonucleotides with a length of at least 100, 150, 200, 250 or 300 nucleotides.

[0016] "Isolated" denotes separated from its natural environment.

[0017] "Polynucleotide" refers in general to polyribonucleotides and polydeoxyribonucleotides, which may be unmodified RNA or DNA or modified RNA or DNA.

[0018] The polynucleotides according to the invention include a polynucleotide according to SEQ ID No. 1 or a fragment produced therefrom, and also polynucleotides that are at least 70% to 80%, preferably at least 81% to 85%, and particularly preferably at least 86% to 90%, and most particularly preferably at least 91%, 93%, 95%, 97% or 99% identical to the polynucleotide according to SEQ ID No. 1 or a fragment produced therefrom.

[0019] The term "polypeptides" is understood to mean peptides or proteins that contain two or more amino acids bound by peptide bonds.

[0020] The polypeptides according to the invention include a polypeptide according to SEQ ID No. 2, in particular those with the biological activity of the sigma factor C and also those that are at least 70% to 80%, preferably at least 81% to 85%, particularly preferably at least 86% to 90%, and most particularly preferably at least 91%, 93%, 95%, 97% or 99% identical to the polypeptide according to SEQ ID No. 2 and that have the aforementioned activity.

[0021] The invention furthermore provides a process for the enzymatic production of amino acids selected from the group L-asparagine, L-threonine, L-serine, L-glutamate, L-glycine, L-alanine, L-cysteine, L-valine, L-methionine, L-isoleucine, L-leucine, L-tyrosine, L-phenylalanine, L-histidine, L-lysine, L-tryptophan and L-arginine, using coryneform bacteria that in particular already produce amino acids and in which the nucleotide sequences coding for the sigC gene are enhanced, in particular overexpressed.

[0022] The term "enhancement" describes in this connection the raising of the intracellular activity of one or more enzymes in a microorganism that are coded by the corresponding DNA, by for example increasing the number of copies of the gene or genes, using a strong promoter, or using a gene that codes for a corresponding enzyme having a high activity, and optionally combining these measures.

[0023] By enhancement measures, in particular overexpression, the activity or concentration of the corresponding protein is in general raised by at least 10%, 25%, 50%, 75%, 100%, 150%, 200%, 300%, 400% or 500%, at most up to 1000% or 2000%, referred to the wild type

protein and/or to the activity or concentration of the protein in the starting microorganism.

[0024] The microorganisms that are the subject of the present invention are able to produce L-amino acids from glucose, sucrose, lactose, fructose, maltose, molasses, starch, cellulose or from glycerol and ethanol. The microorganisms may be representatives of coryneform bacteria, in particular of the genus *Corynebacterium*. In the genus *Corynebacterium* there should in particular be mentioned the species *Corynebacterium glutamicum*, which is known to those skilled in the art for its ability to produce L-amino acids.

[0025] Suitable strains of the genus *Corynebacterium*, in particular of the species *Corynebacterium glutamicum* (*C. glutamicum*), are in particular the known wild type strains

Corynebacterium glutamicum ATCC13032
Corynebacterium acetoglutamicum ATCC15806
Corynebacterium acetoacidophilum ATCC13870
Corynebacterium thermoaminogenes FERM BP-1539
Corynebacterium melassecola ATCC17965
Brevibacterium flavum ATCC14067
Brevibacterium lactofermentum ATCC13869 and
Brevibacterium divaricatum ATCC14020

and L-amino acid-producing mutants or strains produced therefrom.

[0026] The inventors have successfully isolated from *C. glutamicum* the new sigC gene coding for the enzyme sigma factor C.

[0027] In order to isolate the sigC gene or also other genes from *C. glutamicum*, a gene library of this microorganism is first of all incorporated in *Escherichia coli* (*E. coli*). The incorporation of gene libraries is described in generally known textbooks and manuals. As

examples there may be mentioned the textbook by Winnacker: Gene and Klone, Eine Einführung in die Gentechnologie (Verlag Chemie, Weinheim, Germany, 1990 I.B.R.) or the manual by Sambrook et al.: Molecular Cloning, A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1989) I.B.R. A very well-known gene library is that of the E. coli K-12 strain W3110, which was incorporated by Kohara et al. (Cell 50, 495-508 (1987)) I.B.R. into λ vectors. Bathe et al. (Molecular and general genetics, 252:255-265, 1996) I.B.R. describe a gene library of C. glutamicum ATCC13032 that has been incorporated by means of the cosmid vector SuperCos I (Wahl et al., 1987, Proceedings of the National Academy of Sciences USA, 84:2160-2164 I.B.R.) in the E. coli K-12 strain NM554 (Raleigh et al., 1988, Nucleic Acids Research 16:1563-1575 I.B.R.).

[0028] Börmann et al. (Molecular Microbiology 6(3), 317-326) (1992)) I.B.R. in turn describe a gene library of C. glutamicum ATCC13032 using the cosmid pHCT9 (Hohn and Collins, Gene 11, 291-298 (1980) I.B.R.).

[0029] In order to produce a gene library of C. glutamicum in E. coli, there may also be used plasmids such as pBR322 (Bolivar, Life Sciences, 25, 807-818 (1979) I.B.R.) or pUC9 (Vieira et al., 1982, Gene, 19:259-268 I.B.R.). Suitable hosts are in particular those E. coli strains that are restriction-defective and recombinant-defective. An example of such is the strain DH5 α mc^r, which has been described by Grant et al. (Proceedings of the National Academy of Sciences USA, 87 (1990) 4645-4649 I.B.R.). The long DNA fragments cloned with the aid of cosmids can in turn then be subcloned into common vectors suitable for the sequencing and subsequently sequenced, as is described for example by Sanger et al. (Proceedings of the National Academy of Sciences of the United States of America, 74:5463-5467, 1977 I.B.R.).

[0030] The DNA sequences obtained can then be investigated using known algorithms or sequence analysis programs, such as for example that of Staden (Nucleic Acids Research 14, 217-232(1986)) I.B.R., that of Marck (Nucleic Acids Research 16, 1829-1836 (1988)) I.B.R. or the GCG program of Butler (Methods of Biochemical Analysis 39, 74-97 (1998)) I.B.R. Analysis can also be carried out with the "BLAST search program" (Altschul et al., 1997, Nucleic Acids Research, 25:3389-3402)I.B.R., against the non-redundant databank of the "National Center for Biotechnology Information" (NCBI, Bethesda, MD, USA). The relative degree of substitution or mutation in the polynucleotide or amino acid sequence to produce a desired percentage of sequence identity can be established or determined by well-known methods of sequence analysis. These methods are disclosed and demonstrated in Bishop, et al. "DNA & Protein Sequence Analysis (A Practical Approach)", Oxford Univ. Press, Inc. (1997) I.B.R. and by Steinberg, Michael "Protein Structure Prediction" (A Practical Approach), Oxford Univ. Press, Inc. (1997) I.B.R.

[0031] The new DNA sequence of *C. glutamicum* coding for the sigC gene was obtained in this way, and as SEQ ID No. 1 is part of the present invention. The amino acid sequence of the corresponding protein was also derived from the existing DNA sequence using the afore-described methods. The resultant amino acid sequence of the sigC gene product is shown in SEQ ID No. 2.

[0032] Coding DNA sequences that result from SEQ ID No. 1 due to the degeneracy of the genetic code are likewise covered by the present invention. Similarly, DNA sequences that hybridize with SEQ ID No. 1 or parts of SEQ ID No. 1 are also part of the invention. In the specialist field conservative amino acid replacements, such as for example the replacement of glycine by alanine or of aspartic acid by glutamic acid, in proteins are furthermore known as sense mutations that do not lead to any basic change in the

activity of the protein, i.e. are functionally neutral. It is furthermore known that changes at the N-end and/or C-end of a protein do not significantly impair their function or indeed may even stabilize their function. The person skilled in the art can find relevant information on this in, *inter alia*, Ben-Bassat et al. (Journal of Bacteriology 169:751-757 (1987)) I.B.R., in O'Regan et al. (Gene 77:237-251 (1989)) I.B.R., in Sahin-Toth et al. (Protein Sciences 3:240-247 (1994)) I.B.R., in Hochuli et al. (Bio/Technology 6:1321-1325 (1988)) I.B.R. and in known textbooks and manuals on genetics and molecular biology. Amino acid sequences that are obtained in a corresponding manner from SEQ ID No. 2 are likewise covered by the invention.

[0033] In the same way, DNA sequences that hybridize with SEQ ID No. 1 or parts of SEQ ID No. 1 are also covered by the invention. Finally, DNA sequences that are produced by the polymerase chain reaction (PCR) using primers resulting from SEQ ID No. 1, are also part of the invention. Such oligonucleotides typically have a length of at least 15 nucleotides.

[0034] The person skilled in the art can find information on the identification of DNA sequences by means of hybridization in, *inter alia*, the manual "The DIG System User's Guide for Filter Hybridization" published by Boehringer Mannheim GmbH (Mannheim, Germany, 1993) I.B.R. and in Liebl et al. (International Journal of Systematic Bacteriology (1991) 41: 255-260) I.B.R. The hybridization takes place under strict conditions, in other words only hybrids are formed in which the probe and target sequence, i.e. the polynucleotides treated with the probe, are at least 70% identical. It is known that the strictness of the hybridization conditions including the washing step is influenced or determined by varying the buffer composition, temperature and the salt concentration. The hybridization reaction is preferably carried out under conditions that are relatively less strict compared to the wash steps

(Hybaid Hybridisation Guide, Hybaid Limited, Teddington, UK, 1996) I.B.R.

[0035] For the hybridization reaction there may for example be used a 5x SSC buffer at a temperature of ca. 50 - 68°C. In this connection probes can also hybridize with polynucleotides that are less than 70% identical to the probe sequence. Such hybrids are less stable and are removed by washing under stringent conditions. This may be achieved for example by reducing the salt concentration to 2x SSC and then if necessary to 0.5x SSC (The DIG System User's Guide for Filter Hybridisation, Boehringer Mannheim, Mannheim, Germany, 1995 I.B.R.), a temperature of ca. 50 - 68°C being established. It is also possible to reduce the salt concentration down to 0.1x SSC. By stepwise raising of the hybridisation temperature in steps of ca. 1 - 2°C from 50 to 68°C, polynucleotide fragments can be isolated that are for example at least 70% or at least 80% or even at least 90% to 95% identical to the sequence of the probe that is used. Further details relating to hybridisation may be obtained in the form of so-called kits available on the market (e.g. DIG Easy Hyb from Roche Diagnostics GmbH, Mannheim, Germany, Catalog No. 1603558).

[0036] The person skilled in the art can find details on the amplification of DNA sequences by means of the polymerase chain reaction (PCR) in, *inter alia*, the manual by Gait: Oligonucleotides Synthesis: A Practical Approach (IRL Press, Oxford, UK, 1984) I.B.R. and in Newton and Graham: PCR (Spektrum Akademischer Verlag, Heidelberg, Germany, 1994) I.B.R.

[0037] In the course of work carried in connection with the present invention it was established that coryneform bacteria after overexpression of the sigC gene produce amino acids in an improved manner.

[0040] For the enhancement the sigC gene according to the invention was overexpressed for example by means of episomal plasmids. Suitable plasmids are those that are replicated in coryneform bacteria. Numerous known plasmid vectors, such as for example pZ1 (Menkel et al., Applied and Environmental Microbiology (1989) 64: 549-554 I.B.R.), pEKEx1 (Eikmanns et al., Gene 102:93-98 (1991) I.B.R.) or pHS2-1 (Sonnen et al., Gene 107:69-74 (1991) I.B.R.) are based on the cryptic plasmids pHM1519, pBL1 or pGA1. Other plasmid vectors, such as for example those based on pCG4 (US-A 4,489,160 I.B.R.), or pNG2 (Serwold-Davis et al., FEMS Microbiology Letters 66, 119-124 (1990) I.B.R.), or pAG1 (US-A 5,158,891 I.B.R.) may be used in a similar way.

[0041] Furthermore, also suitable are those plasmid vectors with the aid of which the process of gene amplification by integration in the chromosome can be employed, such as has been described for example by Reinscheid et al. (Applied and Environmental Microbiology 60, 126-132 (1994) I.B.R.) for the duplication and amplification of the hom-thrB operon. In this method the complete gene is cloned into a plasmid vector that can replicate in a host (typically E. coli) but not in C. glutamicum. Suitable vectors are for example pSUP301 (Simon et al., Bio/Technology 1, 784-791 (1983) I.B.R.), pK18mob or pK19mob (Schäfer et al., Gene 145, 69-73 (1994) I.B.R.), pGEM-T (Promega Corporation, Madison, WI, USA), pCR2.1-TOPO (Shuman (1994). Journal of Biological Chemistry 269:32678-84 I.B.R.; US-A 5,487,993 I.B.R.), pCR®Blunt (Invitrogen, Groningen, Netherlands; Bernard et al., Journal of Molecular Biology, 234: 534-541 (1993)) I.B.R., pEM1 (Schrumpf et al, 1991, Journal of Bacteriology 173:4510-4516 I.B.R.) or pBGS8 (Spratt et al., 1986, Gene 41: 337-342 I.B.R.). The plasmid vector that contains the gene to be amplified is then transferred by conjugation or transformation into the desired strain of C. glutamicum. The method of conjugation is described for example in Schäfer et al. (Applied and Environmental

Microbiology 60, 756-759 (1994)) I.B.R. Transformation methods are described for example in Thierbach et al. (Applied Microbiology and Biotechnology 29, 356-362 (1988)) I.B.R., Dunican and Shivnan (Bio/Technology 7, 1067-1070 (1989)) I.B.R. and Tauch et al. (FEMS Microbiological Letters 123, 343-347 (1994)) I.B.R. After homologous recombination by means of a crossover event, the resulting strain contains at least two copies of the relevant gene.

[0042] In addition it may be advantageous for the production of L-amino acids to enhance, in particular to overexpress, in addition to the sigC gene also one or more enzymes of the respective biosynthesis pathway, glycolysis, anaplerosis, citric acid cycle, pentose phosphate cycle, amino acid export and optionally regulatory proteins.

[0043] Thus for example, for the production of L-amino acids, in addition to the enhancement of the sigC gene one or more genes selected from the following group may be enhanced, in particular overexpressed:

- the gene dapA coding for dihydrodipicolinate synthase (EP-B 0 197 335 I.B.R.),
- the gene gap coding for glyceraldehyde-3-phosphate dehydrogenase (Eikmanns (1992), Journal of Bacteriology 174:6076-6086 I.B.R.),
- the gene tpi coding for triosephosphate isomerase (Eikmanns (1992), Journal of Bacteriology 174:6076-6086 I.B.R.),
- the gene pgk coding for 3-phosphoglycerate kinase (Eikmanns (1992), Journal of Bacteriology 174:6076-6086 I.B.R.),
- the gene zwf coding for glucose-6-phosphate dehydrogenase (JP-A-09224661 I.B.R.),

- the gene *pyc* coding for pyruvate carboxylase (DE-A-198 31 609 I.B.R.),
- the gene *mgo* coding for malate-quinone-oxidoreductase (Molenaar et al., European Journal of Biochemistry 254, 395-403 (1998) I.B.R.),
- the gene *lysC* coding for a feedback-resistant aspartate kinase (Accession No.P26512; EP-B-0387527 I.B.R.; EP-A-0699759 I.B.R.),
- the gene *lysE* coding for lysine export (DE-A-195 48 222 I.B.R.),
- the gene *hom* coding for homoserine dehydrogenase (EP-A 0131171 I.B.R.),
- the gene *ilvA* coding for threonine dehydratase (Möckel et al., Journal of Bacteriology (1992) 8065-8072) I.B.R.) or the allele *ilvA(Fbr)* coding for a feedback-resistant threonine dehydratase (Möckel et al., (1994) Molecular Microbiology 13: 833-842 I.B.R.),
- the gene *ilvBN* coding for acetohydroxy acid synthase (EP-B 0356739 I.B.R.),
- the gene *ilvD* coding for dihydroxy acid dehydratase (Sahm and Eggeling (1999) Applied and Environmental Microbiology 65: 1973-1979 I.B.R.),
- the gene *zwa1* coding for the Zwa1 protein (DE: 19959328.0 I.B.R., DSM 13115).

[0044] Furthermore, it may be advantageous for the production of L-amino acids, in addition to the enhancement of the *sigC* genes also to attenuate, in particular to reduce, the expression of one or more genes selected from the group

- the gene pck coding for phosphoenol pyruvate carboxykinase (DE 199 50 409.1 I.B.R.; DSM 13047),
- the gene pgi coding for glucose-6-phosphate isomerase (US 09/396,478 I.B.R.; DSM 12969),
- the gene poxB coding for pyruvate oxidase (DE: 1995 1975.7; DSM 13114),
- the gene zwa2 coding for the Zwa2 protein (DE: 19959327.2 I.B.R., DSM 13113).

[0045] The term "attenuation" describes in this connection the reduction or switching off of the intracellular activity of one or more enzymes (proteins) in a microorganism that are coded by the corresponding DNA, by for example using a weak promoter or a gene or allele that codes for a corresponding enzyme having a low activity and/or that inactivates the corresponding gene or enzyme (protein), and optionally combining these measures.

[0046] By means of these attenuation measures the activity or concentration of the corresponding protein is in general reduced to 0 to 75%, 0 to 50%, 0 to 25%, 0 to 10% or 0 to 5% of the activity or concentration of the wild type protein and/or the activity or concentration of the protein in the starting microorganism.

[0047] In addition it may be advantageous for the production of amino acids, in addition to the overexpression of the sigC gene also to switch off undesirable secondary reactions (Nakayama: "Breeding of Amino Acid Producing Micro-organisms", in: Overproduction of Microbial Products, Krumphanzl, Sikyta, Vanek (eds.), Academic Press, London, UK, 1982) I.B.R.

[0048] The microorganisms produced according to the invention are likewise the subject of the invention and may be cultivated continuously or batchwise in a batch process

(batch cultivation) or in a fed batch process (feed process) or repeated fed batch process (repetitive feed process) for the purposes of production of amino acids. A summary of know cultivation methods is given in the textbook by Chmiel (Bioprozeßtechnik 1. Einführung in die Bioverfahrenstechnik (Gustav Fischer Verlag, Stuttgart, 1991)) I.B.R. or in the textbook by Storhas (Bioreaktoren und periphere Einrichtungen (Vieweg Verlag, Brunswick/Wiesbaden, 1994)) I.B.R.

[0049] The culture medium to be used must suitably satisfy the requirements of the relevant strains. Descriptions of culture media for various microorganisms are given in the manual "Manual of Methods for General Bacteriology" of the American Society for Bacteriology (Washington D.C., USA, 1981) I.B.R.

[0050] Carbon sources that may be used included sugars and carbohydrates such as for example glucose, sucrose, lactose, fructose, maltose, molasses, starch and cellulose, oils and fats such as for example soya bean oil, sunflower oil, peanut oil and coconut oil, fatty acids such as for example palmitic acid, stearic acid and linoleic acid, alcohols such as for example glycerol and ethanol, and organic acids such as for example acetic acid. These substances may be used individually or as a mixture.

[0051] Nitrogen sources that may be used include organic nitrogen-containing compounds such as peptones, yeast extract, meat extract, malt extract, corn steep liquor, soya bean flour and urea, or inorganic compounds such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate. The nitrogen sources may be used individually or as a mixture.

[0052] Phosphorus sources that may be used include phosphoric acid, potassium dihydrogen phosphate or dipotassium hydrogen phosphate or the corresponding sodium

salts. The culture medium must furthermore contain salts of metals, such as for example magnesium sulfate or iron sulfate, that are necessary for growth. Finally, essential growth promoters such as amino acids and vitamins may be used in addition to the aforementioned substances. Suitable precursors may furthermore be added to the culture medium. The aforementioned starting substances may be added to the culture in the form of a single one-off batch, or may be suitably metered in during the culture process.

[0053] Basic compounds such as sodium hydroxide, potassium hydroxide, ammonia or ammonia water, or acidic compounds such as phosphoric acid or sulfuric acid, are used in a suitable manner in order to control the pH of the culture. Anti-foaming agents such as for example fatty acid polyglycol esters may be used to control foam formation. In order to maintain the stability of plasmids suitable selectively acting substances such as for example antibiotics may be added to the medium. In order to maintain aerobic conditions, oxygen or oxygen-containing gas mixtures such as for example air are introduced into the culture. The temperature of the culture is normally 20°C to 45°C and preferably 25°C to 40°C. The culture is continued until a maximum of the desired product has been formed. This objective is normally achieved within 10 hours to 160 hours.

[0054] Methods for the determination of L-amino acids are known to the person skilled in the art. The analysis may be carried out for example as described by Spackman et al. (Analytical Chemistry, 30, (1958), 1190) I.B.R. by ion exchange chromatography followed by ninhydrin derivation, or can be carried out by reversed phase HPLC, as described by Lindroth et al. (Analytical Chemistry (1979) 51: 1167-1174) I.B.R.

[0055] The process according to the invention serves for the enzymatic production of amino acids.

[0056] The following microorganisms were filed as a pure culture at the German Collection of Microorganisms and Cell Cultures (DSMZ, Brunswick, Germany) according to the Budapest Convention:

- Escherichia coli DH5 α mc/pEC-XK99EsigCb2ex as DSM 14375 on 29 June 2001
- Corynebacterium glutamicum DSM 5715/pEC-XK99E as DMS 13455 on 17 April 2000.

[0057] The present invention is described in more detail hereinafter with the aid of examples of implementation.

[0058] The isolation of plasmid DNA from Escherichia coli as well as all techniques involved in restriction, Klenow treatment and alkaline phosphatase treatment have been carried out by Sambrook et al. (Molecular Cloning. A Laboratory Manual (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, USA) I.B.R. Methods for the transformation of Escherichia coli are also described in this manual.

[0059] The composition of readily available nutrient media such as LB or TY media are also given in the manual by Sambrook et al.

[0060] Example 1

Production of a genomic cosmid gene library from Corynebacterium glutamicum ATCC 13032

[0061] Chromosomal DNA from Corynebacterium glutamicum ATCC 13032 was isolated as described by Tauch et al. (1995, Plasmid 33:168-179 I.B.R.) and partially cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, product description Sau3AI, Code no. 27-0913-02). The DNA fragments were desphosphorylated with shrimp alkaline phosphatase (Roche Diagnostics GmbH, Mannheim, Germany, product description SAP, Code no. 1758250). The

DNA of the cosmid vector SuperCos1 (Wahl et al. (1987) Proceedings of the National Academy of Sciences USA 84:2160-2164 I.B.R.), obtained from Stratagene (La Jolla, USA, product description SuperCos1 Cosmid Vector Kit, Code no. 251301) was cleaved with the restriction enzyme XbaI (Amersham Pharmacia, Freiburg, Germany, product description XbaI, Code no. 27-0948-02) and likewise dephosphorylated with shrimp alkaline phosphatase.

[0062] The cosmid DNA was then cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, product description BamHI, Code no. 27-0868-04 I.B.R.). The cosmid DNA treated in this way was mixed with the treated ATCC13032-DNA and the batch was treated with T4-DNA ligase (Amersham Pharmacia, Freiburg, Germany, product description T4-DN ligase, Code no. 27-0870-04 I.B.R.). The ligation mixture was then packed into phages using the Gigapack II XL Packing Extracts (Stratagene, La Jolla, USA, product description Gigapack II XL Packing Extract, Code no. 200217 I.B.R.).

[0063] For the infection of the E. coli strain NM554 (Raleigh et al. 1988, Nucleic Acid Research 16:1563-1575 I.B.R.) the cells were taken up in 10 mM MgSO₄ and mixed with an aliquot of the phage suspension. Infection and titration of the cosmid library were carried out as described by Sambrook et al. (1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor) I.B.R., the cells having been plated out on LB agar (Lennox, 1955, Virology, 1:190 I.B.R.) with 100 mg/l ampicillin. Recombinant individual clones were selected after incubation overnight at 37°C.

[0064] Example 2

Isolation and sequencing of the sigC gene

[0065] The cosmid DNA of an individual colony was isolated using the Qiaprep Spin Miniprep Kit (Product No. 27106,

Qiagen, Hilden, Germany) according to the manufacturer's instructions and partially cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, product description Sau3AI, Product No. 27-0913-02). The DNA fragments were dephosphorylated with shrimp alkaline phosphatase (Roche Diagnostics GmbH, Mannheim, Germany, product description SAP, Product No. 1758250). After gel electrophoresis separation, the cosmid fragments were isolated in an order of magnitude of 1500 to 2000 bp using the QiaExII Gel Extraction Kit (Product No. 20021, Qiagen, Hilden, Germany).

[0066] The DNA of the sequencing vector pZero-1, obtained from Invitrogen (Groningen, Netherlands, product description Zero Background Cloning Kit, Product No. K2500-01), was cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, product description BamHI, Product No. 27-0868-04). The ligation of the cosmid fragments in the sequencing vector pZero-1 was carried out as described by Sambrook et al. (1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor I.B.R.), the DNA mixture having been incubated overnight with T4 ligase (Pharmacia Biotech, Freiburg, Germany). This ligation mixture was then electroporated (Tauch et al. 1994, FEMS Microbiol. Letters, 123:343-7 I.B.R.) into the E. coli strain DH5 α MCR (Grant, 1990, Proceedings of the National Academy of Sciences U.S.A., 87:4645-4649 I.B.R.) and plated out onto LB agar (Lennox, 1955, Virology, 1:190 I.B.R.) with 50 mg/l zeocin.

[0067] The plasmid preparation of the recombinant clone was performed with the Biorobot 9600 (Product No. 900200, Qiagen, Hilden, Germany). The sequencing was carried out according to the dideoxy chain termination method of Sanger et al. (1977, Proceedings of the National Academy of Sciences U.S.A., 74:5463-5467 I.B.R.) as modified by Zimmermann et al. (1990, Nucleic Acids Research, 18:1067) I.B.R. The "RR dRhodamin Terminator Cycle Sequencing Kit"

of PE Applied Biosystems (Product No. 403044, Weiterstadt, Germany) was used. The gel electrophoresis separation and analysis of the sequencing reaction was carried out in a "rotiphoresis NF acrylamide/bisacrylamide" gel (29:1) (Product No. A124.1, Roth, Karlsruhe, Germany) using the "ABI Prism 377" sequencing apparatus from PE Applied Biosystems (Weiterstadt, Germany).

[0068] The raw sequencing data obtained were then processed using the Staden program package (1986, Nucleic Acids Research, 14:217-231) I.B.R. Version 97-0. The individual sequences of the pZero1 derivatives were assembled into a coherent contig. The computer-assisted coding region analysis was prepared using the XNIP program (Staden, 1986, Nucleic Acids Research, 14:217-231 I.B.R.).

[0069] The nucleotide sequence obtained is shown in SEQ ID No. 1. The analysis of the nucleotide sequence revealed an open reading frame of 582 base pairs, which was termed the sigC gene. The sigC gene codes for a protein of 193 amino acids.

[0070] Example 3

Production of the shuttle expression vector pEC-XK99EsigCb2ex for the enhancement of the sigC gene in *C. glutamicum*.

[0071] 3.1 Cloning of the sigC gene

Chromosomal DNA was isolated from the strain ATCC 13032 according to the method of Eikmanns et al. (Microbiology 140: 1817-1828 (1994)) I.B.R. The following oligonucleotides for the polymerase chain reaction were selected on the basis of the sequence of the sigC gene known from Example 2 for *C. glutamicum* (see SEQ ID No. 3 and SEQ ID No. 4):

sigCex1:

5' ac ggt acc-ccc tac aca cct tta tgg tg 3' SEQ ID NO: 3

sigCex2:

5' gc tct aga-gtt gac gta gct cat ctg ct 3' SEQ ID NO: 4

[0072] The illustrated primers were synthesised by MWG-Biotech AG (Ebersberg, Germany) and the PCR reaction was carried out according to the standard PCR method of Innis et al. (PCR protocols. A guide to methods and applications, 1990, Academic Press) I.B.R. using Pwo polymerase from Roche Diagnostics GmbH (Mannheim, Germany). With the aid of the polymerase chain reaction the primers permit the amplification of a 667 bp long DNA fragment that carries the sigC gene. Also, the primer sigCex1 carries the sequence for the cleavage site of the restriction endonuclease KpnI, and the primer sigCex2 contains the cleavage site of the restriction endonuclease XbaI, which are underlined in the nucleotide sequence illustrated above.

[0073] The 667 bp long sigC fragment was cleaved with the restriction endonucleases KpnI and XbaI and then isolated from the agarose gel using the QiaExII Gel Extraction Kit (Product No. 20021, Qiagen, Hilden, Germany).

[0074] 3.2 Construction of the shuttle vector pEC-XK99E

The E. coli - C. glutamicum shuttle vector pEC-XK99E was constructed according to the prior art. The vector contains the replication region rep of the plasmid pGA1 including the replication effector per (US-A- 5,175,108 I.B.R.; Nesvera et al., Journal of Bacteriology 179, 1525-1532 (1997)) I.B.R., the kanamycin resistance gene aph(3')-IIa from Escherichia coli (Beck et al. (1982), Gene 19: 327-336 I.B.R.), the replication origin, the trc promoter, the termination regions T1 and T2, the lacI^q gene (repressor of the lac-operon of E.coli) and a multiple cloning site (mcs) (Norrander, J.M. et al. Gene 26, 101-106

(1983) I.B.R.) of the plasmid pTRC99A (Amann et al. (1988), Gene 69: 301-315 I.B.R.).

[0075] The trc promoter can be induced by adding the lactose derivative IPTG (isopropyl- β -D-thiogalactopyranoside).

[0076] The constructed E. coli - C. glutamicum shuttle vector pEC-XK99E was transferred by means of electroporation (Liebl et al., 1989, FEMS Microbiology Letters, 53:299-303 I.B.R.) into C. glutamicum DSM5715. The selection of the transformants was carried out on LBHIS agar consisting of 18.5 g/l brain-heart infusion broth, 0.5 M sorbitol, 5 g/l bacto-tryptone, 2.5 g/l bacto-yeast extract, 5 g/l NaCl and 18 g/l bacto-agar that had been supplemented with 25 mg/l kanamycin. Incubation was carried out for 2 days at 33°C.

[0077] Plasmid DNA was isolated from a transformant by the usual methods (Peters-Wendisch et al., 1998, Microbiology, 144, 915 - 927 I.B.R.), cleaved with the restriction endonuclease HindIII, and the plasmid was checked by subsequent agarose gel electrophoresis.

[0078] The plasmid construct thereby obtained was termed pEC-XK99E (Fig. 1). The strain obtained by electroporation of the plasmid pEC-XK99E into the C. glutamicum strain DSM5715 was identified as DSM5715/pEC-XK99E and filed as DSM13455 in the German Collection of Microorganisms and Cell Cultures (DSMZ, Brunswick, Germany) according to the Budapest Convention.

[0079] 3.3 Cloning of sigC in the E. coli-C. glutamicum shuttle vector pEC-XK99E

The E. coli - C. glutamicum shuttle vector pEC-XK99E described in Example 3.2 was used as vector. DNA of this plasmid was completely cleaved with the restriction enzymes KpnI and XbaI and then dephosphorylated with shrimp

alkaline phosphatase (Roche Diagnostics GmbH, Mannheim, Germany, product description SAP, Product No. 1758250 I.B.R.).

[0080] The ca. 650 bp long sigC fragment described in Example 3.1, which was obtained by PCR and cleaved with the restriction endonucleases KpnI and XbaI, was mixed with the prepared vector pEC-XK99E and the batch was treated with T4-DNA ligase (Amersham Pharmacia, Freiburg, Germany, product description T4-DNA ligase, Code no. 27-0870-04 I.B.R.). The ligation batch was transformed into the E. coli strain DH5 α mc^r (Hanahan, In: DNA cloning. A practical approach. Vol. I. IRL-Press, Oxford, Washington DC, USA I.B.R.). The selection of plasmid-carrying cells was made by plating out the transformation batch on LB agar (Lennox, 1955, Virology, 1:190 I.B.R.) with 50 mg/l kanamycin. After incubation overnight at 37°C recombinant individual clones were selected. Plasmid DNA was isolated from a transformant using the Qiaprep Spin Miniprep Kit (Product No. 27106, Qiagen, Hilden, Germany) according to the manufacturer's instructions and cleaved with the restriction enzymes XbaI and KpnI in order to check the plasmid by subsequent agarose gel electrophoresis. The plasmid obtained was named pEC-XK99EsigCb2ex, and is shown in Fig. 2.

[0081] Example 4

Transformation of the strain DSM5715 with the plasmid pEC-XK99EsigCb2ex

[0082] The strain DSM5715 was transformed with the plasmid pEC-XK99EsigCb2ex using the electroporation method described by Liebl et al., (FEMS Microbiology Letters, 53:299-303 (1989)) I.B.R. The selection of the transformants was carried out on LBHIS agar consisting of 18.5 g/l brain-heart infusion broth, 0.5 M sorbitol, 5 g/l bacto-tryptone, 2.5 g/l bacto-yeast extract, 5 g/l NaCl and 18 g/l bacto-agar that had been supplemented with 25 mg/l kanamycin. Incubation was carried out for 2 days at 33°C.

[0083] Plasmid DNA was isolated from a transformant by the usual methods (Peters-Wendisch et al., 1998, Microbiology, 144, 915 - 927 I.B.R.), cleaved with the restriction endonucleases XbaI and KpnI, and the plasmid was checked by subsequent agarose gel electrophoresis. The strain obtained was named DSM5715/ pEC-XK99EsigCb2ex1.

[0084] Example 5

Production of Lysine

[0085] The *C. glutamicum* strain DSM5715/pEC-XK99EsigCb2ex obtained in Example 4 was cultivated in a nutrient medium suitable for the production of lysine and the lysine concentration in the culture supernatant was determined.

[0086] For this purpose the strain was first of all incubated on an agar plate with the corresponding antibiotic (brain-heart agar with kanamycin (25 mg/l)) for 24 hours at 33°C. A pre-culture was inoculated starting from this agar plate culture (10 ml medium in a 100 ml Erlenmeyer flask). The full medium CgIII was used as medium for the pre-culture.

[0087]

Medium Cg III

NaCl	2.5 g/l
Bacto-Peptone	10 g/l
Bacto-Yeast Extract	10 g/l
Glucose (separately autoclaved)	2% (w/v)

The pH value was adjusted to pH 7.4

Kanamycin (25 mg/l) was added to the medium. The pre-culture was incubated for 16 hours at 33°C and at 240 rpm on a shaker mixer. A main culture was inoculated from this pre-culture so that the initial OD (660 nm) of the main culture was 0.1. The medium MM was used for the main culture.

[0088]

Medium MM

CSL (corn steep liquor)	5 g/l
MOPS (morpholinopropanesulfonic acid)	20 g/l
Glucose (separately autoclaved)	50 g/l
$(\text{NH}_4)_2\text{SO}_4$	25 g/l
KH_2PO_4	0.1 g/l
$\text{MgSO}_4 \cdot 7 \text{ H}_2\text{O}$	1.0 g/l
	10 mg/l
$\text{CaCl}_2 \cdot 2 \text{ H}_2\text{O}$	
$\text{FeSO}_4 \cdot 7 \text{ H}_2\text{O}$	10 mg/l
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	5.0 mg/l
Biotin (sterile filtered)	0.3 mg/l
Thiamine \cdot HCl (sterile filtered)	0.2 mg/l
L-leucine (sterile filtered)	0.1 g/l
CaCO_3	25 g/l

CSL, MOPS and the salt solution were adjusted to pH 7 with ammonia water and autoclaved. The sterile substrate and vitamin solutions as well as the dry autoclaved CaCO_3 were then added.

[0089] The cultivation was carried out in 10 ml volume batches in a 100 ml Erlenmeyer flask equipped with baffles. Kanamycin (25 mg/l) and IPTG (1mM/l) were added. The cultivation was carried out at 33°C and 80% atmospheric humidity.

[0090] After 48 hours the OD was measured at a measurement wavelength of 660 nm using a Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of lysine formed was determined using an amino acid analyzer from Eppendorf-BioTronik (Hamburg, Germany) by ion exchange chromatography and post-column derivation with ninhydrin detection.

[0091] The result of the experiment is shown in Table 1.

Table 1

Strain	OD (660 nm)	Lysine-HCl g/l
DSM5715	11.8	12.99
DSM5715/pEC- XK99EsigCb2ex	12.8	13.96

[0092] This application claims priority to German Priority Document Application No. 100 43 332.4, filed on September 2, 2000 and to German Priority Document Application No. 101 33 426.5, filed on July 10, 2001. Both German Priority Documents are hereby incorporated by reference in their entirety.

SEQUENCING LISTING

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